#### Mutation nomenclature

#### recommendations for the description of DNA changes



http://www.HGVS.org/mutnomen/ HUGO-MDI initiative



## Definitions

# • prevent confusion *mutation*

- change
- disease-causing change

#### polymorphism

- change in >1% population
- not disease causing change

#### • better

neutral terms sequence variant allelic variant alteration CNV (Copy Number Variant) SNV (not SNP)



## Possible variants.

#### • change in sequence

ACATCAGGAGAAGATGTTC GAGACTTTGCCA ACATCAGGAGAAGATGTTT GAGACTTTGCCA ACATCAGGAGAAGATGTTC GAGACTTTGCCA

### • change in amount









Clinical Tools, Inc.

#### Nomenclature







### Mutation nomenclature<sub>2</sub>

#### on behalf of HUGO MDI / HGVS

HUMAN MUTATION 15:7-12 (2000)

#### MDI SPECIAL ARTICLE

#### Mutation Nomenclature Extensions and Suggestions to Describe Complex Mutations: A Discussion

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Consistent gene mutation nomenclature is essential for efficient and accurate reporting, testing, and curation of the growing number of disease mutations and useful polymorphisms being discovered in the human genome. While a codified mutation nomenclature system for simple DNA lesions has now been adopted broadly by the medical genetics community, it is inherently difficult to represent complex mutations in a unified manner. In this article, suggestions are presented for reporting just such complex mutations. Hum Mutat 15:7–12, 2000. © 2000 Wiley-Liss, Inc.

KEY WORDS: complex mutation; mutation detection; mutation database; nomenclature; MDI



#### Nomenclature for the description of sequence variations

(last modified February 12, 2007)

#### Prepared by Johan den Dunnen

#### Contents



 MEW <u>uncertainties in descriptions</u> (incl. arrayCGH, SNP-array, Southern blot data)

#### Current recommendations

- Introduction
- General recommendations
- Specific recommendations
  - ◊ DNA-level

  - Protein-level

#### <u>Checklist</u> - (online help when writing publications)

Example descriptions



#### Follow the recommendations

when you disagree, *start a debate* – do not use private rules, this only causes confusion



#### http://www.HGVS.org/

DNA, RNA, protein

- unique descriptions prevent confusion
- DNA A, G, C, T c.957A>T
- RNA *a, g, c, u r.957a>u*

( deduced mostly )

• protein ( deduced only ) three / one letter amino acid code X = stop codon p.Glu78Gln



Reference Sequence

- use official HGNC gene symbols
- set the residues and numbering NM\_012654.3 : c.957A>T
- provide database reference covering complete sequence largest transcript accession.version number e.g. NM\_012654.3 RefSeq database (curated seqs)



# • indicate type of Reference Sequence

coding DNA	C.
genomic	<b>g</b> .
mitochondrial	т.
RNA	r.
protein	р.



### coding DNA or genomic ?

#### • human genome sequence complete

**COVERS All transcripts** different promoters, splice variants,

different polyA-sites, ...

but

g.21,895,321\_21,895,325del NT\_035218.23 is 70 Mb file new builds follow each other regularly

### • coding DNA

does not cover all variants gives a clue towards position



Residue numbering

• genomic reference sequence from first to last nucleotide 1 to 13,562

not +, - or other signs

### • coding DNA

from first of ATG to last of stop

1 to 963

- (minus) when 5' of ATG incl. 5' of cap site
- \* (asterisk) when 3'of stop codon incl. 3' of polyA-addition site

#### intron

362+1,362+2, ... start ..., 363-2, 363-1 end



### Reference Sequence.



## Residue numbering.

### • **RNA** (deduced mostly)

#### like coding DNA

• protein

( deduced only )

from first to last amino acid 1 to 321



# Types of variation

- simple substitution c.123A>G deletion c.123delA duplication c.123dupA insertion c.123\_124insC other
  - inversion, translocation, transposition
  - complex
  - combinations two alleles >1 per allele

c.[123A>G]+[456C>T] c.[123A>G; 456C>T]



### Substitution

• substitution designated by ">" not used on protein level

#### • examples

*cDNA c.546A>T* (*NM\_012654.3 : c.546A>T*)

genomic g.54786A>G

protein p.GIn78His



## Deletion

- deletion designated by "del" range indicated by "\_"
- examples

c.546delT c.546del

c.586\_591del c.586\_591delTGGTCA or c.586\_591del6

c.781-?\_1392+?del = exon 3 to 6 deletion, breakpoint not sequenced



## Duplication

- duplication designated by "dup" range indicated by "\_"
- examples

c.546dupT c.546dup

c.586\_591dup

c.586\_591dup6 or c.586\_591dupTGGTCA do not describe as insertion

c.781-?\_1392+?dup

= exon 3 to 6 duplication, breakpoint not sequenced



### Insertion

• insertion

designated by "ins" range indicated by "\_" give inserted sequence

#### • examples

**c.546\_547insT** NOT c.546insT

c.1086\_1087insGCGTGA

c.1086\_1087insAB567429.2:g.34\_12,567

when large insert submit to database and give database accession.version number



#### Inversion

- inversion designated by "inv" range indicated by "\_"
- example

c.546\_2031inv



#### Conversion

conversion designated by "con" range indicated by "\_"

• examples

*c.546\_657con917\_1028* 

c.546\_2031conNM\_023541.2:c.549\_2034



#### Translocation

- translocation designated by "t" range indicated by "\_"
- examples

*t*(*X*;4) (*p*21.2;q35) (*c*.857+101\_857+102)



Repeated sequences

 mono-nucleotide stretches g.8932A(18\_23)

> c.345+28T(18\_23) alleles 345+28T[18]+[21]

• di-nucleotide stretches c.7TG(3\_6)

larger g.532\_3886(20\_45) 3.3 Kb repeat



### SNP's

#### • SNP's

clear identifier for each SNP

#### AC043217.2: g.78654 C>G

rs2306220: A>G dbSNP entry

DXS1219: g.117CA(18\_26) alleles g.117CA[20]+[24]



## Specific codes

- codes used
  - +, -,

>

.

,

"

- substitution
- range

\*

- more changes in one allele
  - more transcripts / mosaicism
- () uncertain
- [] allele
- del deletion
- dup duplication
- ins insertion
- inv inversion
- con conversion
- ext extension
- X stop codon
- fsX frame shift
- o opposite strand
- t translocation



Changes in 2 alleles

- recessive disease report combination of changes
  - allele indicated by "[]" separated by "+"
- examples

c.[546C>T]+[1398deIT] or [c.546C>T]+[c.1398deIT] c.[546C>T]+[?] c.[546C>T]+[=]



## Alleles

- recessive disease
   c.[546C>T]+[2398delT]
   c.[546C>T]+[?]
- more changes in 1 allele c.[546C>T; 2398delT]
- alleles unknown c.[246C>T(+)2398deIT] parents not analysed
- more variants from 1 allele

mosaicism - c.[=, 546C>T] two transcripts - r.[=, 512\_636del]



### Frame shifts

• short form *p.Arg*83fs

(sufficient)

Iong from

(more detail)

p.Arg83SerfsX15

do not try to include changes at DNA level

indicate

first amino acid changed position first changed amino acid length shifted frame (from first changed to X incl.) do not describe del, dup, ins, etc.



# Complex

#### • deletion / insertions

"indel"

c.1166\_1177delinsAGT

#### • descriptions nay become complex

only an expert understands the "code" consider database submission

description: AC111747.1



### Publications

#### Published

patient#	protein	DNA	Remarks							
QL43.2	Tyr151Asp	451T>C	parents unrelated							
 QL43.2	 frame shift	 976delA	 parents unrelated							
Correct										

DNA	RNA	protein	Remarks		
451T>C(+) 976delA]	? or NA	? (p.[Tyr151Asp (+)Pha326fal)	parents unrelated		
	451T>C(+)	451T>C(+) ?	451T>C(+) ? ?		



#### Problematic

#### • coding DNA Reference Sequence

#### no reference to genomic sequence so, c.5477-137A>G ?

#### no reference to intron numbering so, c.IVS20-1G>A ?

new exon identified (CFTR, SMN1) exons 12, 13, 13A, 14 exons -2, -1, 1, 2, 3, ...

difference with genome browser always from 1 to end



Gene structure ?

• how are exons numbered ? often confusing

• how are introns numbered ?

SMN1 exon 7 **exon 0 / intron 0** where is exon 0 in a gene ? ..., -2, -1, 0, 1, 2, ... or ..., -2, -1, 1, 2, ...

#### >> changes difficult to find non-expert, student, ...



### Exon 0



Baker, Nature 442: 916





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Progranulin Aug.2006

#### **Problematic**<sub>2</sub>

#### • not described at DNA level

e.g. Abstract / Title / Results gives p.Tyr151Asp should give c.451T>C or c.451T>C (p.Tyr151Asp) at least on first appearance

#### • range in description

c.635+12\_14del seems clear (probably c.635+12\_635+14del) but c.35+121\_128del ?



**Problematic**,

#### • insertions

503insT is not clear ins at position 503 or after position 503 ? what about c.591-3insT ?

#### • from name to one-letter AA-code

G? - Glu, Gly, Gln A? - Ala, Arg, Asn, Asp Phe - P? (no F!)

#### • assume most 3' residue is changed

ATGTC AAAAA TCGG c.10delA versus c.6delA



Problematic.

- recessive disease combination of alleles not listed
- AA numbering leader sequence not included
- reporting "polymorphisms" p.Arg123Arg

useless and equivocal (no info, 5 possibilities DNA) 36A/G

unclear, c.36A>G or p.Ala36Gly ?

• experimental proof ? "change affects splicing" was RNA actually analysed ?



#### Problematic.

#### • change detected

 DNA
 c.2873G>T

 RNA
 r.2843\_2873del

 protein
 Arg945fsX23

#### • list this as ?

substitution, splice mutation or frame shift



This meeting

(seen at this meeting)

- XYZ\_e01(-2599)
- -45A>C ... -3I/D ... 86C>G
- XYZ-267 ... +10 ... +80 (+10 positioned 5' of 5' UTR)
- A196G
- IVS0+5G>C



#### One debate, ... of many

reported: c.451T>C / p.Tyr151Asp text states: no RNA of this allele

so correct: c.451T>C (r.?/p.0)

*author refuses to change, editor accepts* 

what enters the database ?

is p.Tyr151Asp deleterious ?

pathogenic change may be elsewhere other DNA change giving similar protein change



# Why bother ?

Lydon, April 12, 2008—a XBG patient and his parents sued the department of clinical diagnosis in Lydon, the XBG mutation database, and the journal Human Mutation. The complaint was that serious and culpable mistakes were made during the clinical diagnosis of the pregnancy in the XBGfamily, that ultimately led to the birth of an affected child. A paper published in Human Mutation listed the sequence variant detected in the family as "nonpathogenic." Careful examination would have revealed that the change was clearly pathogenic (a nonsense mutation). However, the accused parties failed to verify the data of the original report and just copied it. 

HUMAN MUTATION 22:181-182 (2003)



# Mutalyzer

#### reference sequence input: GenBank or private

### output: correctly described change

DNA, protein, all annotated transcripts, RE-sites, ...

<sup>©</sup>Peter Taschner

Mutalyzer Menu	Mutalyzer - Sequence variant nomenclature check
<u>Start Page</u>	The Mutalyzer interface comes in a few flavors. http://www.LOVD.nl/mutalyzer/
Name Generator	First of all there is the standard interface. This interface gives you user readable output. Go to <u>Mutalyzer -Name Generator</u>
Name Checker	Secondly you have an option to check whether your own given mutation name is correct or not. Go to <u>Mutalyzer -Name Checker</u>
SNP Converter	Thirdly you have the option to do a batch query/name check. For this, you will need an tab delimited
Batch Checker	text file, which you can generate yourself, or which you can generate with the page provided. Go to <u>Mutalyzer -Batch Name Checker</u>
<u>GenBank Uploader</u>	Last of all you have the option to upload your own Sequences. For this, you will need a text file in GenBank format.
<u>Disclaimer</u>	Go to <u>Mutalyzer -GenBank Uploader</u>
Questions: <u>mutalyzer@humqen.nl</u>	The Engine of Mutalyzer is based on Python. The Python Scripting Language was chosen for good readability and therefore easy maintenance of the engine which can be done by most people.
	MC © 2006 LUMC ? Help   Disclaimer

### Generates ref.seq.

			3.03			3.0				•			3.03			3.5				• 3	
GA	GGC	CAA	GCT	ACT	GCG	TCA	ACA	CAA	AGG	CCG	CCT	GGA	AGC	CAG	GAT	GCA	AAT	CCT	GGA	A	10740
Ε	A	K	L	L	R	Q	H	K	G	R	L	E	A	R	M	Q	I	L	Е		3580
										•									1	<u>76</u> .	
GA	CCA	CAA	TAA	ACA	GCT	GGA	GTC	ACA	GTT	ACA	CAG	GCT	AAG	GCA	GCT	GCT	GGA	GCA	A	CCC	10800
D	H	Ν	K	Q	L	E	S	Q	L	H	R	L	R	Q	L	L	E	Q	1	P	3600
										•										• 3	
CA	GGC	AGA	GGC	CAA	AGT	GAA	TGG	CAC	AAC	GGT	GTC	CTC	TCC	TTC	TAC	CTC	TCT	ACA	GAG	·G	10860
Q	A	Ε	A	K	v	Ν	G	Т	Т	v	s	S	Ρ	s	Т	s	L	Q	R		3620
			5			55				•											
TC	CGA	CAG	CAG	TCA	GCC	TAT	GCT	GCT	CCG	AGT	GGT	TGG	CAG	TCA	Ai						

SDSSQPMLLRVVGSQ: Nyetn

GI EEDLLSPPQDTST (

GAGCAACTCAACAACTCCTTCCCTAGTTCAAGAG | GAAGAAATI E Q L N N S F P S S R G | R N

#### **Dystrophin gene - Intron 14**

(intronic numbering for cDNA Reference Sequence)

aaaagtaaagatttatgtttatttattccttggaattctttaatgtcttgcag 1705-1

agteettagtateagteatgacaga**tga**agaaggageagaa<u>taa</u>a:

gattcccgcatggtttttataatattcatacaacaaagaggatta

Ivo Fokkema / Johan den Dunnen

### Mutation database

# Submit <u>all</u> the changes you have, NOW

(without errors)





Recent suggestion

#### • Copy Number Variants

(last-present\_first-deleted)\_(last-deleted\_first-present)del

BAC / PAC probe derived (AC123322.10\_AL109609.5)\_(AL451144.5\_AL050305.9)del AL109609.5\_AL451144.5del where non-del ?

g.(32,218,983\_32,238,146)\_(32,984,039\_33,252,615)del NCBI build 36.1

SNP-array

(rs2342234\_rs3929856)\_(rs10507342\_rs947283)del

g.(32,218,983\_32,238,146)\_(32,984,039\_33,252,615)del NCBI build 36.1

